

Role of peptide valency on PEG-b-PPS micelles for integrin receptor blocking and anti-angiogenesis

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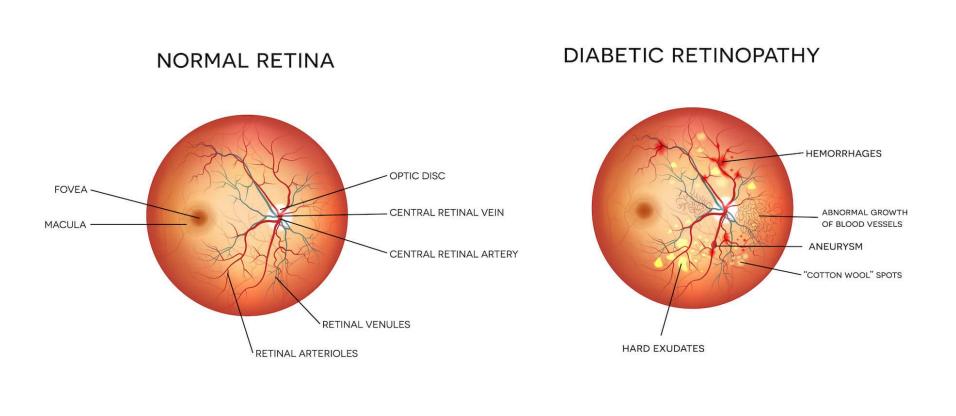
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INTRODUCTION

Anti-integrin therapy was recently established as a promising strategy for angiogenesis-related disease, including Diabetic Retinopathy (DR), Retinopathy of (ROP), Age-related Prematurity degeneration (AMD) [1],[2].

- It was previously reported that PEG-b-PPS micelles bound anti-integrin peptide Serine-Aspartic acid-(SDV) demonstrated enhanced angiogenesis at lower peptide concentration [3].
- However, optimization of the surface density of peptides is imperative to improve integrin clustering and subsequent cell signaling.
- The abnormal growth of blood vessels in the eye can be destroyed by cell death or apoptosis.
- Hence for triggering apoptosis, an optimum valency of peptides must be presented on the surface of the micelles to bind to the ανβ3 integrin and block it.



Liquid Chromatography-Mass Spectrometry (LC-MS)

Fig 1: LC-MS analysis. Graph showing LC-MS analysis with the purity of

the peptide samples. The peak obtained on the chromatogram co-relates

to the molecular weight of the peptide. (a) SDV peptide has a mol wt of

Cell viability assay

SGV-5 SDV-1 SDV-3 SDV-5 cRGD-5 Blank MC

Fig 3: Cell viability analysis using MTT assay. Graph showing cell viability of

HUVEC cells after 24 h incubation with micelles of different peptide densities,

blank micelles compared cRGD bound micelles and control endothelial cells.

1350g/mol, **(b)** SGV- 1291 g/mol, **(c)** cRGD-1174 g/mol

density increases.

p<0.05 is considered as significant

Particle size and Zeta potential

HYPOTHESIS

We **hypothesize** that incorporating an optimum density of anti-integrin peptides on micelles will significantly increase the efficiency of anti-angiogenesis.

OBJECTIVES

The **objective** of this study is to synthesize and characterize PEG-b-PPS micelles with a different surface density of anti-integrin peptides and study the angiogenic ability using primary human umbilical vein endothelial cells (HUVEC).

C: Expression of integrin ανβ3 **B: Characterization** A(ii): Preparation of PEG-b-PPS-anti **A(i): Peptide Modification** αvβ3 peptide micelles Anti-ανβ3 peptide-PEG-Palmitoleic acid Injected to the RP-HPLC at 42 and 45 retention time F: In vitro tube formation assay

12,000 cells/well was

seeded in a 96-

well plate

coated with

Matrigel incubated for

and hour at 37°C

Once confluent

it was treated

with anti-αvβ3

peptide samples of

varying surface

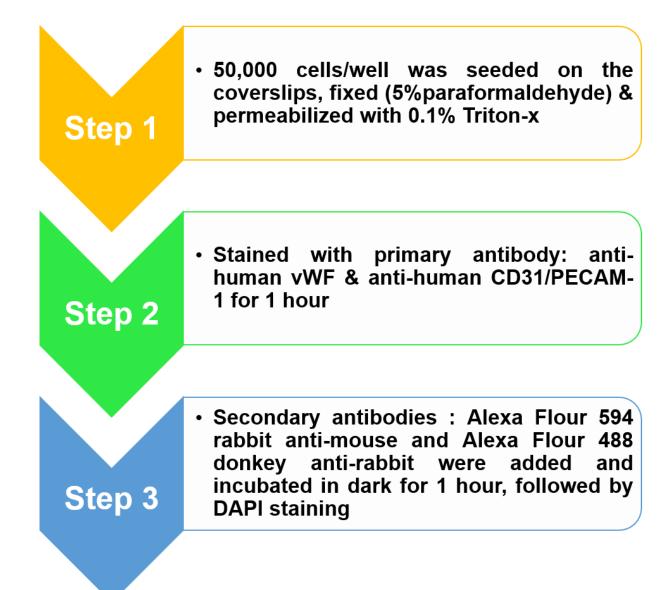
densities

Allowed to form

tubes for 4hrs

by incubating it

E: Cell viability D: Expression of angiogenic markers

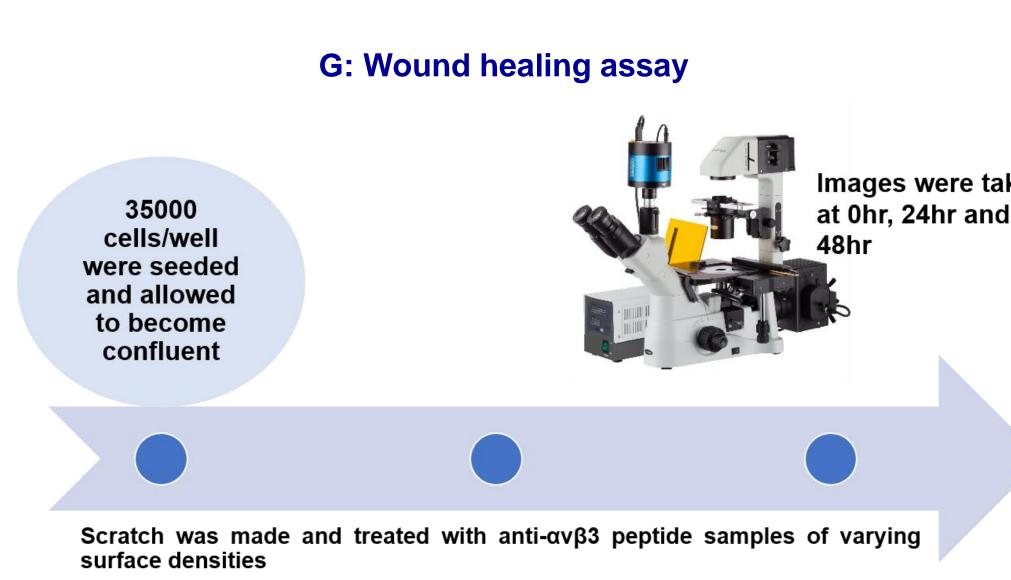


Samples were confirmed using

LC-MS was

performed

seeding density of 8000 - 10,000 Treated HUVEC's with varying surface densities of anti-ανβ3



quantified by ImageJ and the

intensity was

RESULTS

Expression of angiogenic markers

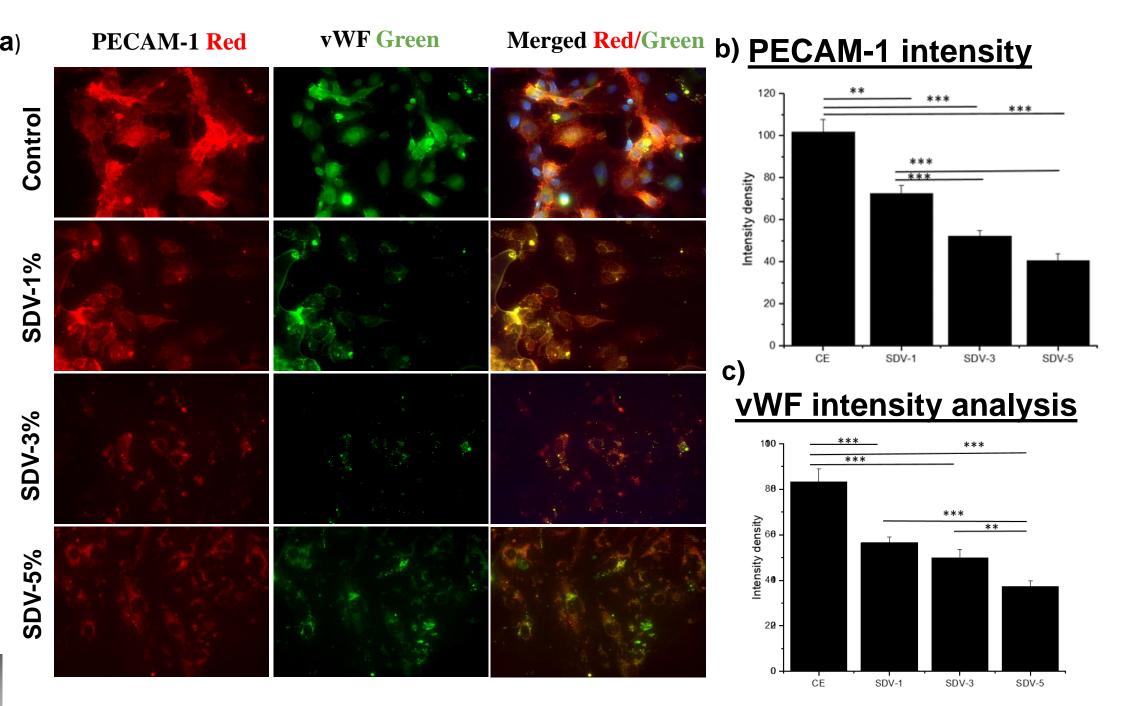


Fig 4:a) Immunostaining with angiogenic markers Platelet Endothelial Cell Adhesion molecule and von Wilebrand factor (vWF-red) showing the decrease in intensity from control to SDV-5%. Quantitative expression of b) PECAM-1 and c) vWF

Expression of integrin ανβ3

ncubated for 4 hours

MATERIALS & METHODS

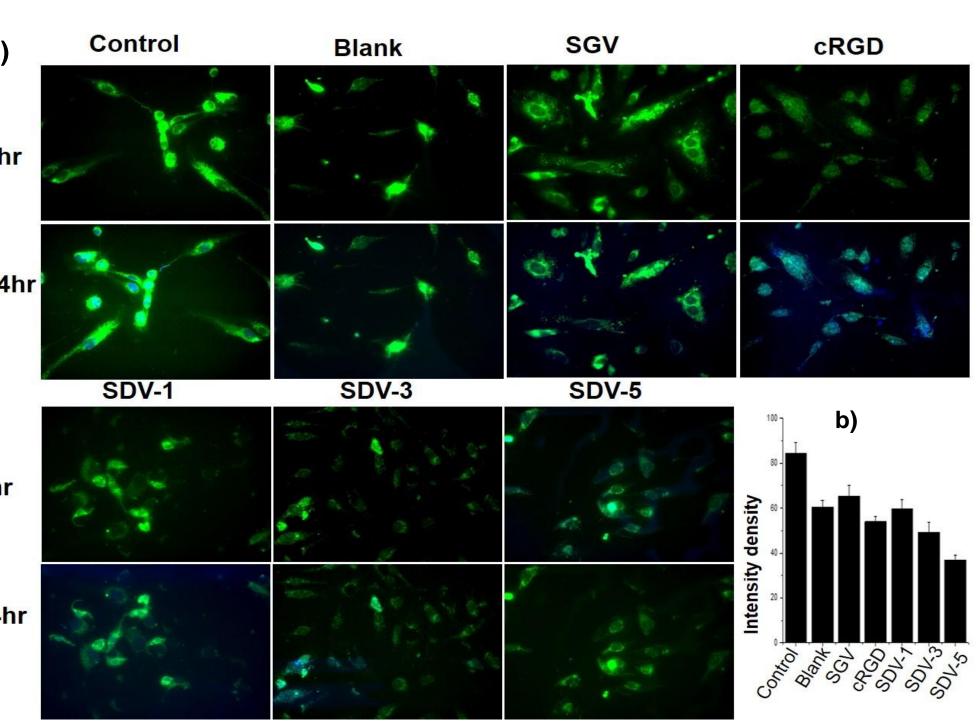


Fig 6: a) Immunostaining with Anti-human CD51/CD61 (Integrin $\alpha v\beta 3$) showing the inhibition of integrin $\alpha v\beta 3$ by SDV peptides with varying surface densities and comparing it scrambled peptide and cRGD and b) quantitative expression of Integrin αvβ3

Invitro tube formation assay

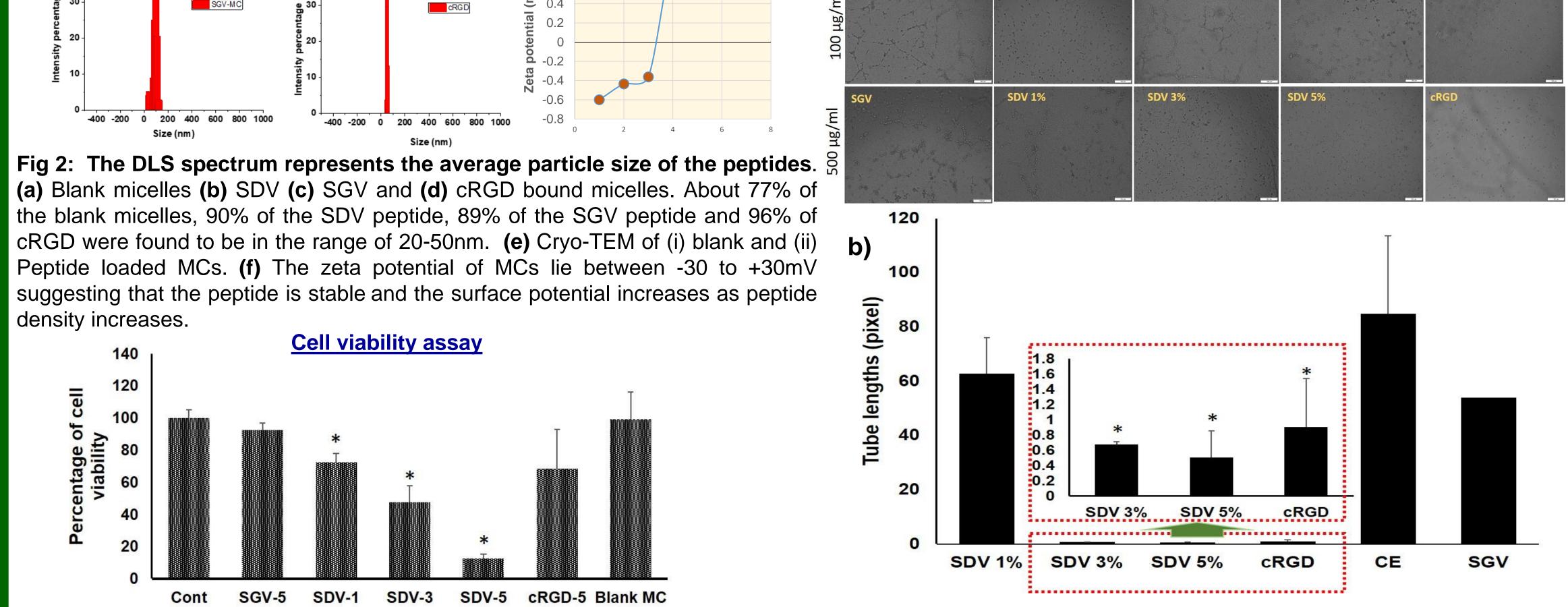


Fig 5: a) The in vitro angiogenesis was inhibited in the HUVEC cells by the PEG-b-PPS- anti αvβ3 peptides in a dose dependent manner and b) the tube lengths were analyzed using anti-angiogenesis analyzer plugin Image J. p<0.05 is considered as significant

Wound-healing assay

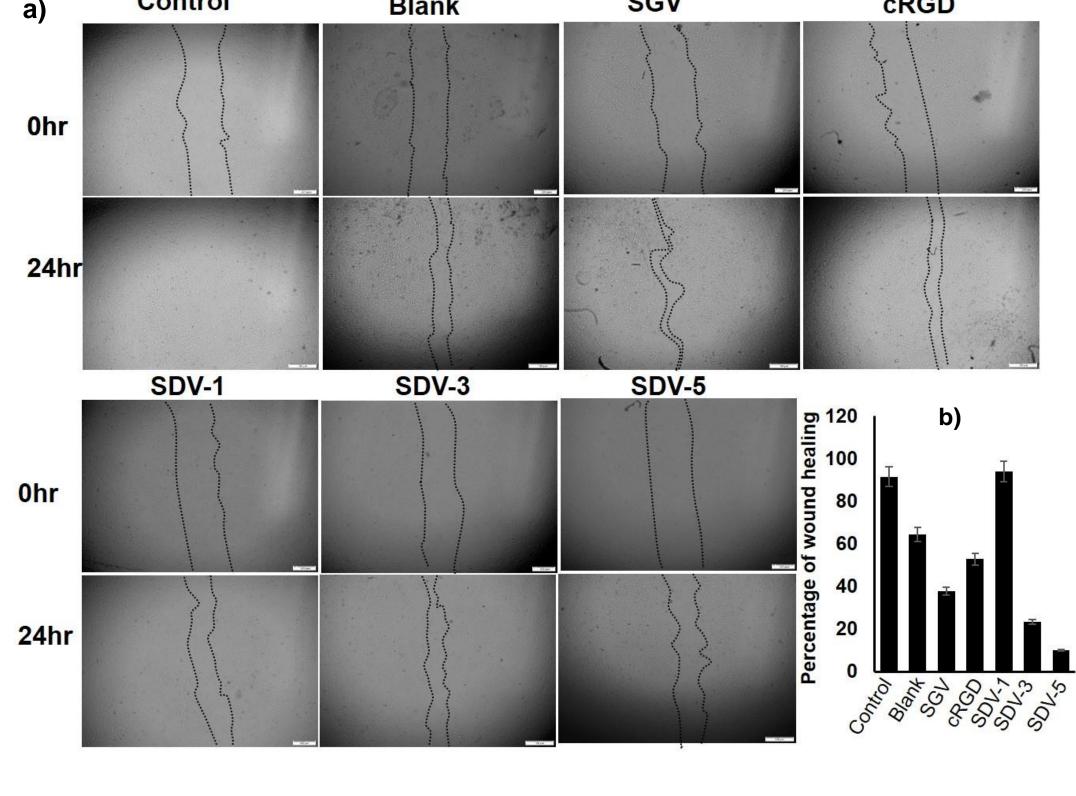


Fig 7: a)The inhibitory effect of the PEG-b-PPS anti ανβ3 peptides on migration of HUVEC's after 24hrs of treatment and b) the %of wound healing analyzed using ImageJ software. SDV-5% showed maximum inhibition than SDV-3% and SDV-1%.

DISCUSSION

- LC-MS determined the purity of the peptide samples and the DLS confirmed the particle size of the peptides to be between 20-50nm.
- The expression of angiogenic markers vWF which is responsible for the formation of blood vessels and PECAM-1 which is responsible for vascular integrity confirmed that the HUVEC's used in our experiment have angiogenic properties. A significant increase in the inhibition of vWF and PECAM-1 was also observed when the surface density of the peptides was increased from 1% to 5%.
- Similarly with the increase in the peptide density from SDV-1% to SDV-5%, the increase in the inhibition of integrin ανβ3 was observed. This led to the decrease in the intensity of the anti-human CD51/CD61 stained cells.
- At 500µg/ml concentration, SDV-1, 3, and 5 induced significantly higher toxicity (27.71±5.43%, 52.54±10.48%, 87.27±2.70%, respectively) compared to control. However, cRGD-MC showed approximately 40% toxicity at both concentrations studied, whereas random peptide SGV-MC did not show significant toxicity at any concentrations studied.
- SDV-5 showed inhibition of tube formation when compared to SDV-3 and SDV-1 suggesting that better anti-angiogenic effect can be expected from SDV-5%.
- Similarly, the migration of the HUVEC's in the wound healing assay saw maximum inhibition by SDV-5% than SDV-3 and SDV-1.
- However, one major limitation of this study is that the efficiency of the peptide to inhibit angiogenesis is analyzed in vitro but, the effect of the peptide in vivo to inhibit angiogenesis is yet to be studied.

FUTURE DIRECTION

- To develop an efficient PEG-b-PPS –anti ανβ3 peptide for sustained release and to bring about an effective antiangiogenic response for patients suffering from diseases like diabetic retinopathy (DR).
- Perform in vivo studies using diabetic retinopathy animal model to confirm the sustained release kinetics, tissue distribution and anti-angiogenic efficiency.

CONCLUSIONS

- > From this study, it can be concluded that peptide density on the micellar surface is crucial in enhancing integrin clustering for anti-angiogenesis.
- > The study provided a detailed understanding of the development of a novel ligand targeted therapeutic strategy, which can be utilized for various disease conditions.

REFERENCES

1.Gibson et al., 2012, American Journal of Preventive Disease 2. Cao et al., 2018, Cell Communication and Signaling 3. Nagaraj *et al.*, 2020, Nanomaterials

ACKNOWLEDGMENTS





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